

Review Article

The Role of Cardiokines in Heart Diseases: Beneficial or Detrimental?

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Cardiovascular disease remains the leading cause of morbidity and mortality, imposing a major disease burden worldwide. Therefore, there is an urgent need to identify new therapeutic targets. Recently, the concept that the heart acts as a secretory organ has attracted increasing attention. Proteins secreted by the heart are called cardiokines, and they play a critical physiological role in maintaining heart homeostasis or responding to myocardial damage and thereby influence the development of heart diseases. Given the critical role of cardiokines in heart disease, they might represent a promising therapeutic target. This review will focus on several cardiokines and discuss their roles in the pathogenesis of heart diseases and as potential therapeutics.

1. Introduction

Despite an obvious decrease in the number of deaths attributable to cardiovascular disease (CVD) during the preceding decades, it remains the primary killer worldwide and, unfortunately, the hospitalization rate in patients less than 55 years old has not been improved [1]. With changing lifestyles and an aging population, cardiovascular risk factors have become more prevalent, and the number of people living with CVD is increasing, thereby causing a seemingly unbearable economic burden for society [2]. It is therefore necessary to identify new strategies to achieve more accurate diagnosis, which could result in better treatments for CVD.

Cardiac myocytes have been reported to synthesize and release natriuretic peptides [3–5]. In addition to its role as a mechanically beating organ, the role of the heart as a secretory organ has attracted increasing attention. It has been well recognized that investigating the pathogenesis of heart failure (HF) has transformed from an investigation of cardiac hemodynamics to neuroendocrinological assessments. Cardiac dysfunction can significantly activate the natriuretic peptide system [4], and atrial natriuretic peptide (ANP) and brain natriuretic peptide (BNP) are both closely related to

the motion of the cardiac wall (such as under conditions of excessive blood, assumption of the trendelenburg position, or increased central venous pressure) and improving the signal transduction between the heart and peripheral organs. Given that this paracrine/autocrine signaling within the heart plays a critical physiological role in the process of cardiac diseases, there is an urgent need to identify novel therapeutic targets based on the secretory function of the heart.

A growing body of evidence showed that the peptides or proteins secreted from cardiac cells can be considered cardiokines [6]. Most cardiokines, as important mediators, play pivotal roles in maintaining healthy heart homeostasis or in the response to myocardial damage. It has been reported that cardiokines not only have physiological involvement in the stress response, damage repair, and myocardial remodeling, but could also participate in protein synthesis in distal organ tissues and systemic metabolic processes [7, 8]. Additionally, cardiokines are differentially expressed in various physiological conditions of the heart, and these secreted cardiokines are intended to maintain healthy cardiac function through paracrine/autocrine pathways or affect the response of cardiomyocytes and cardiac fibroblasts (CFs) to pathological abnormalities caused by heart damage or

other associated inflammatory processes, ultimately eliciting a protective or harmful effect on cardiac function [7, 8].

Many researchers have realized that cardiokines could act as biomarkers to evaluate cardiac function, and therefore contribute to clinical diagnosis, and provide novel therapeutic targets for cardiac diseases. Increasing attention has been paid by researchers in this field to identifying novel cardiokines, with a view to understanding abnormalities in intercellular communication to better diagnose heart disease. In addition to regular laboratory examinations, advanced techniques including gene expression analysis, array screening, cloning, and other methods provide advanced approaches to identify novel cardiokines and determine the networks between cardiokines that are dysregulated during cardiac stress [7, 8]. In this review, we briefly introduce several cardiokines and discuss their roles in the pathogenesis and treatment of cardiac diseases. Furthermore, we summarize the physiological effects of these cardiokines in cardiac diseases in Table 1.

2. The Beneficial Role of Cardiokines in CVD

2.1. Natriuretic Peptides. Natriuretic peptides, and in particular ANP and BNP, secreted by the cardiovascular system, have a particularly large impact on the occurrence and development of CVD in a paracrine/autocrine manner [9, 10]. It is well recognized that ANP and BNP are useful for the clinical diagnosis, treatment, and prognosis of CVD [9, 10]. There is evidence that ANP is significantly elevated in patients with left ventricular dysfunction which is independent of clinical symptoms, and that the ANP levels in the circulation are negatively correlated with ejection fraction (EF) [11]. Interestingly, increased levels of ANP in the circulation are positively correlated with the severity of congestive heart failure (CHF), whereas ANP levels are significantly decreased after an improvement in CHF symptoms.

BNP, also known as B-type natriuretic peptide, is mainly secreted by ventricular myocytes [12]. Although BNP has a variety of biological actions, cardiomyocytes only directly synthesize the precursor of BNP (the 108 amino acid proBNP) [12, 13]. ProBNP, which is initially stored in cardiomyocytes, is released and instantly decomposes into BNP and inactive NT-proBNP in equimolar quantities when the ventricular walls experience stretching forces or ventricular pressure is increased [3]. It therefore appears that BNP and its precursor play a clinically significant role in response to various CVDs such as HF, hypertension, and arrhythmias [14, 15]. In addition, BNP contributes to better diagnosis of acute HF, and in particular HF classification [16]. Similarly, BNP is closely associated with the prognosis of chronic HF and also could be used as an independent prognostic marker for CVD. The European Society of Cardiology has recommended BNP as an indicator for the diagnosis of HF in 2001, and the 2005 American guidelines for HF further reinforced this recommendation [17]. Theoretically, BNP and NT-proBNP are equally significant for CVD diagnosis. A recent systematic review suggested that BNP strongly correlates with NT-proBNP, and joint measurements could improve the accuracy and reliability of the diagnosis of acute or chronic HF [18]. Compared with BNP, NT-proBNP possesses a longer half-life

and is more stable in plasma. NT-proBNP levels are closely related to newly synthesized rather than stored BNP and NT-proBNP preferentially reflects the activation of the BNP pathway [19].

2.2. Interleukin-33. The inflammatory response is thought to be one of the most important mechanisms in the process of atherosclerosis. Abnormalities in the levels of various inflammatory cytokines have been found in patients with acute coronary syndrome [20, 21]. Extremely elevated interleukin (IL) levels in the heart and myocardial necrosis during acute myocardial infarction (MI) indicate that ILs act as an important regulatory factor in acute MI [22].

IL-33, mainly secreted by CFs, is a paracrine signaling molecule involved in crosstalk between fibroblasts and cardiomyocytes, and it is also the specific ligand for soluble ST2 (sST2), which is confirmed to be a cardiomyocyte protein. Mechanical traction and stimulation remarkably upregulate the expression of IL-33 in cardiomyocytes and fibroblasts, as well as levels of ST2 (sST2 levels are significantly higher than those of ST2), and then sST2 exhibits competitive inhibition, thereby blocking the IL-33/ST2 signaling pathway and attenuating the protective effect of IL-33 on cardiomyocyte hypertrophy and myocardial fibrosis [23, 24]. Furthermore, an aldosterone receptor antagonist indirectly upregulates IL-33 expression by reducing ST2 levels, enhancing the IL-33/ST2 signaling pathway and then reducing inflammation and fibrosis after MI [25]. In addition, serum ST2 levels are closely associated with the prognosis of MI and HF [26], and they have been recommended as a biomarker for additional risk stratification in the American Heart Failure Guidelines 2013 [27]. Studies of coronary heart disease patients also show that genetic polymorphisms of these inflammatory cardiokines could increase the risk of coronary heart disease [28, 29].

2.3. Follistatin. Follistatin is an extracellular modulator that selectively binds to proteins of the transforming growth factor- β super family (TGF- β ; discussed later). Follistatin-like 1 (FSTL1), also known as transforming growth factor β -stimulated clone 36 (TSC-36) [30], has been identified as a cardioprotective factor that could protect cardiomyocytes and decrease apoptosis induced by ischemia/reperfusion (IR) injury [31, 32]. The physiological mechanisms underlying FSTL1 action are quite different from those of other follistatins. A recent study revealed that the expression of FSTL1 in the ischemic area after MI is obviously increased in fibroblasts, but not in cardiomyocytes [33]. Compared with wild-type mice, the activation and differentiation of myofibroblasts in FSTL1 gene-knockout mice were attenuated, and the accelerated formation of extracellular matrix (ECM), such as collagen and fibrous proteins, in the ischemic heart was obviously reduced, suggesting an increased risk of mortality with cardiac rupture. These findings indicate that FSTL1 could stimulate the activation of fibroblasts and protect against cardiac rupture and left ventricular remodeling [34]. Interestingly, Altekoster and colleagues reported that bioengineered FSTL1 patches reduce heart scarring and induce angiogenesis, which may provide an effective strategy

TABLE 1: Summary of the physiological roles of cardiokines in cardiac diseases.

| Cardiokine | Beneficial or detrimental | Action mechanisms | Dose-response | Types of cardiac diseases | Predictor |
|----------------------------|------------------------------|---|---------------|---------------------------|-----------|
| Natriuretic peptide | | | | | |
| ANP [11] | Beneficial | - | Yes | HF | - |
| BNP [3, 12-15] | Beneficial | - | Yes | HF | Yes |
| Interleukin | | | | | |
| IL-33 [23-27] | Beneficial | IL-33/ST2, sST2 | - | HF, MI, CH, MF | Yes (ST2) |
| IL-6 [95-97] | Detrimental | gpl30 | Yes | MI | Yes |
| IL-18 [98, 99] | Detrimental | - | - | CAHD | - |
| IL-1 β [91-94] | Detrimental | NO synthase | - | CH, ACS | - |
| Follistatin | | | | | |
| FSTL1 [30-38] | Beneficial | AMPK, BMP-4 | - | MI, CH, HF | - |
| FSTL3 [131, 132] | Detrimental | - | - | CH | - |
| FGF | | | | | |
| FGF21 [39-45] | Beneficial | β -Klotho, ERK | - | CAHD | - |
| FGF23 [109-113] | Harmful | ERK1/2 | - | MF, CH | - |
| Sfrp | | | | | |
| Sfrp-3 [46-50] | Beneficial | Wnt signaling | Yes | HF, MI | - |
| Sfrp-2 [51-54] | Protective | Wnt signaling, BMP-1/Tolloid-like metalloproteinase | - | MI | - |
| MIF [55-59] | | | | | |
| NRG [60-62] | Beneficial (the cardiogenic) | - | - | MI | - |
| ADM [63-69] | Beneficial | - | - | MI, MF | - |
| PI16 [70, 71] | Beneficial | - | Yes | MI, CAHD, MI | Yes |
| Neurotrophins | | | | | |
| MANF [72-74] | Beneficial | KLF2, ERK5 | - | CH, HF | - |
| CDNF [75] | Beneficial | ATF6 | - | MI, CH | - |
| NDNF [76] | Beneficial | - | - | MI | - |
| BDNF [77-81] | Beneficial | Focal adhesion kinase/Akt | - | MI | - |
| Ang-II [82-86] | Detrimental | TrkB | - | MI | - |
| TNF- α [100-108] | Detrimental | TRPM7, AT1R, JNK, ERK | - | ME, HF | - |
| MMPs [114-117] | Detrimental | PLA2/AA, PKA, Cx40 | Yes | HF, ACS, Arrhythmia | - |
| | | TIMP s | - | HF, CAHD, ACS | - |

TABLE 1: Continued.

| Cardiokine | Beneficial or detrimental | Action mechanisms | Dose-response | Types of cardiac diseases | Predictor |
|----------------------------|---------------------------|------------------------------------|---------------|---------------------------|-----------|
| PDGF [118–123] | Detrimental | PDGFR- α and PDGFR- β | - | ME, CAHD | - |
| TGF- β [34, 124–135] | Undetermined | TGF- β receptor 1/2 | - | MI | - |
| CTRP9 [135–142] | Undetermined | gCTRP9, AdipoRI, AMPK, Akt | - | MI, HF, CH | - |

ACS: acute coronary syndrome; ADM: adrenomedullin; Ang-II: angiotensin-II; AMPK: adenosine 5'-monophosphate-activated protein kinase; ANP: atrial natriuretic peptide; AT1R: Ang-II 1 receptor; ATF6: activating transcription factor 6; BDNF: brain-derived neurotrophic factor; Bmp1: bone morphogenic protein 1; BNP: brain natriuretic peptide; CAHD: coronary atherosclerotic heart disease; CDNF: cerebral dopamine neurotrophic factor; CH: cardiac hypertrophy; CTRP9: C1q/TNF-related protein 9; Cx40: connexin 40; ERK: extracellular regulated protein kinases; FGF: fibroblast growth factor; FSTLI: follistatin-like 1; GDF-15: growth differentiation factor-15; gp130: glycoprotein 130; HF: heart failure; IL: interleukin; JNK: c-Jun N-terminal kinase; MANF: mesoscopic astrocyte-like neurotrophic factor; MF: myocardial fibrosis; MI: myocardial infarction; MIF: macrophage migration inhibitory factor; MMPs: matrix metalloproteinases; NDNF: neuron-derived neurotrophic factor; NO: nitric oxide; PDGF: platelet-derived growth factor; P116: protease inhibitor 16; PKA: protein kinase A; PLA2/AA: phospholipase A2/arachidonic acid; Sfrp-3: secreted frizzled-related protein-3; TGF- β : transforming growth factor- β ; TIMP: tissue inhibitor of metalloproteinase; TNF- α : tumor necrosis factor- α ; TRPM7: transient receptor potential melastatin-7 channels; TSC-36: transforming growth factor β -stimulated clone 36.

for reducing the risk of cardiac rupture and unfavorable remodeling after MI [35].

In a study by Tanaka et al. [36, 37], FSTL1 expression induced by cardiac stress was described to modulate cardiac hypertrophy, while FSTL1 knockout mice showed more serious cardiac hypertrophy and cardiac dysfunction after HF. Similarly, Ogura and coworkers [38] demonstrated that recombinant FSTL1 administered in mice or pig models could remarkably reduce the proportion of the MI region after IR, subsequently inhibiting apoptosis and the inflammatory response via adenosine 5'-monophosphate- (AMP-) activated protein kinase and bone morphogenetic protein-4-dependent mechanisms. Moreover, overexpression of FSTL1 also minimized the deleterious effects of IR injury [38]. All these findings indicate that FSTL1 may become a therapeutic target for cardiac hypertrophy or other heart diseases.

2.4. Fibroblast Growth Factor 21. Fibroblast growth factors (FGFs) play a definite role in inducing angiogenesis, repairing impaired endothelial cells, and promoting vascular smooth muscle cell proliferation [39, 40]. To date, there are 22 identified human or murine FGFs. FGF sequences between different animals have a high relative homology. FGFs transmit signals inside cells through their related external receptors on the cell membrane [41].

FGF21, a new member of the FGF family, consists of 209 amino acids. It is physiologically decomposed into 181 amino acids for maturity. FGF21 is mainly produced and released from the liver, but a recent study indicates that cardiomyocytes and cardiac microvascular endothelial cells (CMECs) might also express FGF21 to improve cardiac remodeling and reduce heart damage [42]. FGF21 specifically binds to FGFR1c in the heart to exert biological effects through activation of its coreceptor β -klotho [41]. The N-terminus of FGF21 binds to FGFR1c while the C-terminus binds to β -Klotho with a high affinity to form a complex that phosphorylates FGFR1c and activates downstream signal transduction (extracellular regulated protein kinases (ERK) signaling pathway), which explains the intracellular effects of FGF21 [43, 44]. In addition, FGF21 is secreted into the systemic circulation by damaged myocardial or endothelial cells and affects cell surface receptors to regulate lipid metabolism and protect against oxidative stress or inflammatory injury, thereby leading to amelioration of atherosclerosis and protection against ischemic myocardium and IR injury [45].

2.5. Secreted Frizzled-Related Protein. Secreted frizzled-related protein-3 (Sfrp-3) is the strongest Wnt signal antagonist in the Sfrps family. Recent studies revealed that Wnt signaling is involved in cardiac hypertrophy [46, 47], as well as in the progression of late atherosclerosis and it is significantly associated with vascular inflammation, endothelial dysfunction, calcification, and intimal thickening [48]. Askevold et al. [49] demonstrated that Sfrp3 levels are markedly elevated in patients with HF, which is consistent with the New York Heart Association (NYHA) functional classification. In contrast, Sfrp3 is significantly decreased in end-stage HF patients after treatment [50]. It is therefore likely that Wnt/Sfrp3 might play a significant role in left

ventricular remodeling. Furthermore, mRNA levels of Sfrp3 are elevated in cardiomyocytes, endothelial cells, and CFs in the MI region in mouse models [49, 50]. Sfrp3 appears to be a biomarker reflecting the pathogenesis of HF and a potential therapeutic target for heart diseases.

Similar to Sfrp3, Sfrp2 plays a critical role in inhibiting apoptosis and inflammatory reactions through interfering with Wnt signaling [51]. The expression of Sfrp2 in the murine heart after MI is significantly upregulated, suggesting that Sfrp2 is a stress-inducible cardiokine [52]. Furthermore, an exogenous injection of Sfrp2 at a therapeutic dose could deactivate the activity of bone morphogenetic protein 1 (Bmp1)/Tolloid-like metalloproteinase [53], thereby inhibiting collagen deposition at the late stage of MI and improving left ventricular function. Therefore, Sfrp2 is a potential therapeutic target for antifibrosis therapies [54].

2.6. Macrophage Migration Inhibitory Factor. Macrophage migration inhibitory factor (MIF) is a highly conserved factor closely related to the inflammatory response. MIF is released from necrotic cardiomyocytes after MI, and the levels in circulation are rapidly increased after stimulation since the cells produce and store MIF before the inflammatory response [55, 56]. Furthermore, increased MIF levels are closely associated with the infarct region [57], making it a potential biomarker. A previous study reported that the -173G/C polymorphism of MIF is associated with coronary heart diseases [58].

After MI, MIFs of different cellular sources play opposing physiological roles. In a study by White et al. [59], mice with MIF deficiency in bone marrow derived-cells had a lower incidence of cardiac rupture after MI, whereas MIF deficiency in somatic/cardiac cells accelerated ventricular dilatation and dysfunction. In conclusion, the majority of MIFs in the infarcted myocardium are from infiltrating inflammatory cells, rather than cardiogenic cells. Inhibiting MIF from inflammatory cells could protect cardiac function and improve MI prognosis. Moreover, MIF protects the heart from short-term hypoxia, but, with the prolongation of ischemia and hypoxia, the protective effect of MIF in the heart is gradually weakened. Meanwhile, the proinflammatory effect of MIF gradually emerges, eventually exacerbating myocardial injury [55]. Collectively, as mentioned previously, this bidirectional effect of MIF may be associated with its different origins.

2.7. Neuregulin. Neuregulin (NRG) is a member of the epidermal growth factor (EGF) family and is mainly secreted by microvascular endothelial cells and endocardium in the heart. NRG can promote angiogenesis, reverse myocardial remodeling, and improve apoptosis and oxidative stress. It has recently been reported that NRG is also an important signaling protein in the cardiovascular system and in regulating cardiac development and cardiac function, since the tyrosine kinase receptor of NRG (ErbB) has been detected on the surface of cardiomyocytes [60]. Hedhli et al. demonstrated that hypoxia-reoxygenation could induce myocardial endothelial cells to express and release NRG, and that NRG could protect adult mouse cardiomyocytes against apoptosis during hypoxia-reoxygenation [61]. In addition, NRG may

directly improve fibrosis [62] and induce the production and secretion of IL-1 α and repair factors (like crypto-1), which affect cardiac healing through paracrine signaling [60]. These findings indicate that endothelium-derived NRG has a protective effect in the ischemic myocardium and it may represent a new therapeutic target for heart diseases.

2.8. Adrenomedullin. Adrenomedullin (ADM) is a product of vascular endothelial cells, smooth muscle cells, and cardiomyocytes and is thought to be a local factor in controlling vascular tension, cardiac contractility, and renal sodium excretion [63]. Cheung et al. suggested a significant increase of plasma ADM levels in patients with CHF because of neuroendocrine reactions [64].

ADM levels are associated with endothelial injury and can indicate the severity of atherosclerotic endothelial cell injury [65]. In addition, a previous study showed that ADM is beneficial for HF and MI and that short-term treatment using ADM reduces the area of MI and IR injuries because of its antioxidant and antiapoptosis effects [66]. A follow-up study by Nishida et al. demonstrated that a high risk for CVD was associated with abnormal plasma levels of ADM in 121 patients [67]. This study suggested that plasma ADM is an independent predictor of cardiovascular events in high risk patients [68, 69]. In conclusion, ADM is a predictive biomarker for the onset of CVD, and in particular HF.

2.9. Protease Inhibitor 16. Protease inhibitor 16 (PI16) is a protein secreted by cardiomyocytes and it might elicit inhibitory effects on myocardial hypertrophy. It is strongly upregulated in the early phase of HF and restrains the growth of cardiomyocytes in vitro and in vivo. Overexpression of PI16 inhibits hypertrophy in cultured cardiomyocytes. In contrast, PI16 knockout activates hypertrophy [70]. Interestingly, it has been determined that the Kruppel-like factor 2- (KLF2-) mediated ERK5-dependent signaling pathway may be involved in PI16 inhibition of endothelial migration and proliferation, which contributes to the maintenance of vascular homeostasis [71]. Adenovirus-mediated PI16 overexpression inhibits the secretion of matrix metalloproteinases (MMPs; discussed later). In addition, inflammatory cytokines including IL-1 β and tumor necrosis factor- α (TNF- α) have a significant impact on the NF- κ B signaling pathway by strongly inhibiting the expression of PI16. It is therefore likely that PI16 is an endogenous protective cardiokine, which could be applied as a therapeutic target in heart diseases, such as HF and hypertrophic cardiomyopathy.

2.10. Neurotrophins. Mesoscopic astrocyte-like neurotrophic factor (MANF), an endoplasmic reticulum (ER) stress protein secreted by cardiomyocytes, protects cells against stress in a paracrine/autocrine manner. ER stress could activate transcription factor 6 (ATF6), which induces the expression and secretion of MANF [72]. It has also been reported that MI activates MANF expression in cardiomyocytes and non-myocytes, attenuating cardiac hypertrophy and myocardial ischemic injury. Additionally, a preclinical study showed that MANF knockout increases myocardial ischemia after

ischemia reperfusion [72]. In contrast, increasing the level of MANF by administering recombinant MANF protein protects against heart injury in mice [73]. An in vitro study using cultured cardiomyocytes indicated that MANF could exert inhibitory effects on stress-induced hypertrophy [74]. It is therefore likely that MANF is a beneficial cardiokine that assists with recovery following cardiac diseases.

Similar to MANF, cerebral dopamine neurotrophic factor (CDNF) is an ER stress protein that can be induced by the activation of ER stress in cultured cardiomyocytes [75]. In addition, overexpression of CDFN improves cell viability and protects cardiomyocytes against ER stress-induced apoptosis.

Neuron-derived neurotrophic factor (NDNF) [76], a proangiogenic secreted protein with a fibronectin type III domain, improves poor myocardial remodeling after infarction and enhances cardiac cell survival and angiogenesis through a focal adhesion kinase/Akt-dependent pathway. Mice intramuscularly injected with adenovirus vectors expressing NDNF exhibit better left ventricular systolic and diastolic function after MI, as well as enhanced capillary formation and reduced posterior cardiomyocyte apoptosis and hypertrophy. Consequently, treatment of cultured cardiomyocytes using recombinant NDNF could reduce apoptosis under hypoxic conditions.

Brain-derived neurotrophic factor (BDNF) is widely expressed in many nonneural tissues such as vascular endothelial cells and myocardial cells, and it plays a role in regulating vascular repair and promoting wound healing [77]. It plays an important role in the improvement of myocardial microcirculation after myocardial damage [78]. Myocardial ischemia preconditioning increases the expression of BDNF mRNA and protein in cardiomyocytes, suggesting that BDNF exerts a protective action against myocardial IR by reducing apoptosis and enhancing antioxidant activity in the heart [79]. Tropomyosin receptor kinase B (TrkB), a functional receptor of BDNF, mediates downstream signaling through dimerization with BDNF and intracellular kinase-specific tyrosine phosphorylation, thereby enhancing cardiac microvascular endothelial cell proliferation and survival [80, 81]. It is therefore possible that BDNF is helpful during myocardial repair by promoting neovascularization.

3. The Detrimental Role of Cardiokines in CVD

3.1. Angiotensin-II. Angiotensin-II (Ang-II) is mainly synthesized and released by the renin-angiotensin-aldosterone system (RAAS) [82]. Interestingly, a study by Chen et al. demonstrated that Ang-II could also be produced by cardiomyocytes and fibroblasts in the heart, which elicits biological effects through paracrine or autocrine pathways [83]. CFs are the key cells initiating the formation of myocardial fibrosis. Zhang et al. demonstrated that Ang-II has the potential to abnormally increase the growth of CFs, resulting in myocardial fibrosis, through a transient receptor potential melastatin-7 channel-mediated (TRPM7, calcium channels) inward calcium current [84]. In addition, Ang-II promotes the expression of the Ets-1 gene in CFs, which is involved in tissue fibrosis remodeling, in a time and concentration

dependent manner via the Ang-II 1 receptor (AT1R), c-Jun N-terminal kinase (JNK), or ERK signaling pathway [85]. Furthermore, pretreatments using losartan (an AT1R inhibitor), PD98059 (an ERK inhibitor), or SP600125 (a JNK inhibitor) facilitate the inhibition of cell proliferation and myocardial fibrosis by significantly downregulating profibrogenic factors such as connective tissue growth factor (CTGF) and plasminogen activator inhibitor-1 (PAI-1) [86]. Similarly, Ang-II leads to cardiac diastolic dysfunction by inducing myocardial fatty acid oxidation [87]. Given the critical role of Ang-II in CHF, it may aid accurate diagnosis and act as a predictor for clinical outcomes [88, 89]. Moreover, NT-proBNP is highly associated with altered levels of Ang-II. Together, this evidence suggests that combined measurements of NT-proBNP with Ang-II could effectively improve diagnostic accuracy for CHF [90].

3.2. Interleukin-1 β , Interleukin-6, and Interleukin-18. In contrast to the role of IL-33, some interleukins have a detrimental effect in heart diseases. A preliminary study indicated that IL-1 β might contribute to the onset of cardiomyocyte hypertrophy [91] and that sustained high levels of IL-1 β not only cause cardiac pump impairment but also aggravate undesirable cardiac remodeling [92]. Additionally, IL-1 β induces the expression of nitric oxide (NO) synthase and weakens the positive effects of β -adrenergic agonists on cardiomyocytes [93, 94]. Furthermore, the level of IL-6 in the blood is elevated in patients with MI, and sustained excess IL-6 production leads to cardiac damage through glycoprotein 130 (gp130) [95, 96]. Circulating levels of IL-6 are also closely related to the severity of left ventricular dysfunction and are an effective predictor for subsequent clinical complications [97]. Moreover, IL-18 is an independent risk factor in the formation and development of plaques in atherosclerosis by reducing the stability of atherosclerotic plaques and ECM degradation [98, 99].

3.3. Tumor Necrosis Factor- α . Tumor necrosis factor- α (TNF- α) is expressed by myocardial cells under stress and it is a harmful cardiokine involved in atherosclerosis [100]. TNF- α is upregulated during CHF and it contributes to impaired myocardial contractility, cardiomyocyte apoptosis, and myocardial remodeling. More importantly, serum levels of TNF- α are associated with CHF severity [101, 102].

There are several studies on the relationship between the expression of TNF- α and IL-1 β with secondary ventricular arrhythmias in patients with acute coronary syndromes [103, 104], in which TNF- α and IL-1 β are significantly upregulated and the levels increase with the deterioration of ventricular arrhythmia. Consequently, TNF- α and IL-1 β are helpful in predicting the occurrence of secondary ventricular arrhythmia in patients with acute coronary syndrome and could be applied as useful biomarkers in estimating the severity of ventricular arrhythmia. Three possibilities underlie these pathological mechanisms: (1) TNF- α may be related to the opening of calcium ion channels in cardiomyocytes through a signal transduction pathway such as phospholipase A2/arachidonic acid (PLA2/AA), which affects cardiomyocyte repolarization and impairs contraction [105, 106]; (2)

TNF- α could alter the potassium channels of cardiomyocytes via a protein kinase A (PKA) signaling pathway and inhibit rectifying potassium currents, ultimately causing myocardial abnormalities [107]; (3) TNF- α has also been shown to downregulate the expression of connexin 40 (Cx40) in gap junctions, thereby affecting intercellular communication and inducing arrhythmias [108].

3.4. Fibroblast Growth Factor 23. As a member of the FGF family, FGF23 derived from injured myocardial tissues, in contrast with the beneficial role of FGF21, promotes fibrosis and diastolic dysfunction after MI or IR [109]. In this pathological process, FGF23 is frequently accompanied by the activation of β -Klotho and TGF- β [110]. Recombinant FGF23 administration can directly induce pathological cardiac hypertrophy [111]. Furthermore, FGF23 elevation in the circulation is highly associated with an increased risk of cardiovascular events, such as myocardial ischemia, stroke, and cardiovascular disease-related deaths [112]. Intriguingly, the ERK1/2 pathway plays a critical role in FGF23 function and could enhance phosphate-mediated vascular calcification by promoting osteoblastic differentiation [113].

3.5. Matrix Metalloproteinases. Matrix metalloproteinases (MMPs) are a group of proteins that are capable of selectively degrading ECM and regulate most of the ECM remodeling in CHF patients via cardiac remodeling and left ventricular dilatation [114]. All MMPs are negatively regulated by tissue inhibitors of metalloproteinase (TIMPs), and MMP/TIMP imbalance may result in heart disease [115].

MMPs are dramatically increased during HF progress and recovery [116]. In sheep models simulating the process of left ventricular hypertrophy, failure, and recovery, different MMP subtypes and their TIMP inhibitors were abnormally regulated during the process of myocardial ECM remodeling, thereby affecting the development of HF and ventricular remodeling [117]. In addition, the levels of MMP-2 and MMP-9 in patients with coronary atherosclerotic heart disease are significantly increased, while exogenous inhibitors restrain the expression and activity of MMPs to maintain the stability of atherosclerotic platelets [117]. Together, this evidence indicates that MMPs are harmful cardiokines, which exacerbate the prognosis of heart disease. TIMPs may act as new therapeutic targets for cardiac diseases via inhibition of MMPs, but this approach requires further investigation.

3.6. Platelet-Derived Growth Factors. Platelet-derived growth factors (PDGFs) are commonly expressed in the myocardium and interstitial fibroblasts [118]. PDGFs stimulate pathological hyperplasia of fibroblasts and convert them into myofibroblasts by activating specific receptors (PDGFR- α and PDGFR- β), resulting in the production of a significant amount of collagen, which is involved in the development of fibrosis [119, 120]. PDGF is also closely related to the occurrence and progression of coronary atherosclerosis, in which PDGF-D affects the stability of coronary atherosclerotic plaques [121]. A previous study indicated that PDGF could promote the accumulation of smooth muscle cells and the formation of foam cells in atherosclerosis [122]. In

addition, PDGF has the potential to induce the division and proliferation of damaged epithelial or endothelial cells, resulting in the subsequent aggravation of atherosclerosis [123].

4. Undetermined Role of Cardiokines in CVD

4.1. Transforming Growth Factor- β . Transforming growth factor- β (TGF- β) is secreted by multiple cell types including cardiomyocytes, which affects the regulation of cell growth and differentiation. It has been reported that TGF- β is a regulator of neutrophil infiltration and can clean up the necrotic cells and matrix caused by the inflammatory response during MI recovery. TGF- β is therefore significantly associated with undesirable ventricular remodeling after infarction [124–126]. Recently, accumulating evidence has shown that members of the TGF- β superfamily including growth differentiation factor-15 (GDF-15), myostatin, and activin A play a cardioprotective role in maintaining normal cardiac homeostasis and inhibiting myocardial hypertrophy [34, 127–129]. In addition, FSTL1 (mentioned earlier in the article) is a regulatory factor for GDF-15 in cultured cardiac myocytes or mouse hearts, where it protects against cardiac stress [130], while overexpression of FSTL3 can reverse the effects of activin A to promote cardiomyocyte survival and inhibit cardiomyocyte hypertrophy [131, 132], suggesting that activin A and FSTL3 exert counteractive effects in regulating myocardial cell growth and fibrosis and the response to ischemic stress. Collectively, these findings deepen our understanding of cardiokine networks.

In contrast, several studies have demonstrated that TGF- β inhibition in the late stage of MI may improve myocardial remodeling, while TGF- β inhibition in the early stage has opposite effects [133, 134], suggesting that the activation of TGF- β may protect the heart from ischemic injury at an early stage, but its beneficial effect is later diminished. Similarly, Rainer et al., using a mouse model of selective TGF- β receptor 1 or 2 knockout, demonstrated that the inhibitory effects of TGF- β signaling on cardiomyocytes could significantly suppress neutrophil aggregation in the heart, restrain the inflammatory reaction, prevent cardiac rupture after MI, and improve remodeling [126]. Selective TGF- β inhibition improves ventricular remodeling by directly reducing the production of proinflammatory cytokines/chemokines and inhibiting neutrophil activation and migration through inducing the synthesis of other protective cardiokines. However, specific and pleiotropic characteristics of TGF- β may contribute to various potential side-effects of nonselective inhibition [135]. The adverse roles of TGF- β at different stages of MI could also increase its complexity for clinic application, so careful assessments must be undertaken to determine how TGF- β could be adopted in clinical treatment to improve ventricular remodeling.

4.2. C1q/TNF-Related Protein 9. C1q/TNF-related protein 9 (CTRP9) is a novel cardiokine with high relative homology to adiponectin (APN) [136], which is primarily secreted by adipose tissue and cardiac endothelial cells [137]. CTRP9 can maintain homeostasis and improve the prognosis of

heart disease via its inhibitory effects on inflammation, post-IR injury, and ventricular remodeling after MI [138–141]. Kambara et al. [138, 142] demonstrated that CTRP9 levels are reduced by nearly 50% in mice following myocardial IR injury with an increase of blood levels of free fatty acid (FFA). An exogenous injection of CTRP9 could dramatically attenuate MI and improve cardiomyocyte apoptosis after myocardial ischemia, suggesting that CTRP9 is a protective cardiokine in the cardiovascular system. In contrast, it was also found that heterozygous or homozygous CTRP9 knockout mice showed less cardiac hypertrophy and pulmonary congestion after pressure overload, as well as better systolic cardiac function, compared with wild-type mice [137]. Moreover, CTRP9 overexpression induces serious cardiac dysfunction, suggesting that CTRP9 might exert a deleterious effect on cardiac hypertrophy and HF. Since the role of CTRP9 in cardiac diseases has not been fully determined, future studies on clarifying how CTRP9 triggers different signaling pathways during the pathological process of cardiac hypertrophy are required.

5. Conclusions

The physiological role of cardiokines has been attracting more attention since cardiokines have shown significant potential as biomarkers to evaluate cardiac function and as therapeutic targets for cardiac diseases. It has been suggested not only that cardiokines have physiological effects on cardiac tissues, but that they may also exert regulatory effects on peripheral organs and tissues [143, 144]. Further detailed studies on the role of cardiokines in the crosstalk between the heart and peripheral organs are required. In addition, the regulatory effects of cardiokines are often complex, as they can exert bidirectional actions to promote the repair of cardiac injury and/or aggravate an imbalance of cardiac function. Since the physiological role of cardiokines in cardiac diseases is not fully determined, additional studies are warranted.

Abbreviations

| | |
|---------|---|
| ACS: | Acute coronary syndrome |
| ADM: | Adrenomedullin |
| Ang-II: | Angiotensin-II |
| AMPK: | Adenosine 5'-monophosphate-activated protein kinase |
| ANP: | Atrial natriuretic peptide |
| APN: | Adiponectin |
| AT1R: | Ang-II 1 receptor |
| ATF6: | Activating transcription factor 6 |
| BDNF: | Brain-derived neurotrophic factor |
| Bmp1: | Bone morphogenic protein 1 |
| BNP: | Brain natriuretic peptide |
| CAHD: | Coronary atherosclerotic heart disease |
| CDNF: | Cerebral dopamine neurotrophic factor |
| CFs: | Cardiac fibroblasts |
| CH: | Cardiac hypertrophy |
| CHF: | Congestive heart failure |

| | |
|-----------------|--|
| CMEC: | Cardiac microvascular endothelial cells |
| CTGF: | Connective tissue growth factor |
| CTRP9: | C1q/TNF-related protein 9 |
| CVD: | Cardiovascular disease |
| Cx40: | Connexin 40 |
| ECM: | Extracellular matrix |
| EF: | Ejection fraction |
| EGF: | Epidermal growth factor |
| ER: | Endoplasmic reticulum |
| ERK: | Extracellular regulated protein kinases |
| FFA: | Free fatty acid |
| FGF: | Fibroblast growth factor |
| FSTL1: | Follistatin-like 1 |
| GDF-15: | Growth differentiation factor-15 |
| gp130: | Glycoprotein 130 |
| HF: | Heart failure |
| IL: | Interleukin |
| IR: | Ischemia/reperfusion |
| JNK: | c-Jun N-terminal kinase |
| MANF: | Mesoscopic astrocyte-like neurotrophic factor |
| MF: | Myocardial fibrosis |
| MI: | Myocardial infarction |
| MIF: | Macrophage migration inhibitory factor |
| MMPs: | Matrix metalloproteinases |
| NDNF: | Neuron-derived neurotrophic factor |
| NO: | Nitric oxide |
| NRG: | Neuregulin |
| NYHA: | New York Heart Association |
| PAI-1: | Plasminogen activator inhibitor-1 |
| PDGF: | Platelet-derived growth factor |
| PII6: | Protease inhibitor 16 |
| PKA: | Protein kinase A |
| PLA2/AA: | Phospholipase A2/arachidonic acid |
| RAAS: | Renin-angiotensin-aldosterone system |
| Sfrp-3: | Secreted frizzled-related protein-3 |
| TGF- β : | Transforming growth factor- β |
| TIMP: | Tissue inhibitor of metalloproteinase |
| TNF- α : | Tumor necrosis factor- α |
| TRPM7: | Transient receptor potential melastatin-7 channels |
| TSC-36: | Transforming growth factor β -stimulated clone 36. |

Conflicts of Interest

The authors have no conflicts of interest to disclose.

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References

- [1] K. A. Wilmot, M. O'Flaherty, S. Capewell, E. S. Ford, and V. Vaccarino, "Coronary heart disease mortality declines in the United States from 1979 through 2011: Evidence for stagnation in young adults, especially women," *Circulation*, vol. 132, no. 11, pp. 997–1002, 2015.
- [2] G. Nicholson, S. R. Gandra, R. J. Halbert, A. Richhariya, and R. J. Nordyke, "Patient-level costs of major cardiovascular conditions: A review of the international literature," *ClinicoEconomics and Outcomes Research*, vol. 8, pp. 495–506, 2016.
- [3] T. Sudoh, K. Kangawa, N. Minamino, and H. Matsuo, "A new natriuretic peptide in porcine brain," *Nature*, vol. 332, no. 6159, pp. 78–81, 1988.
- [4] A. J. de Bold, H. B. Borenstein, A. T. Veress, and H. Sonnenberg, "A rapid and potent natriuretic response to intravenous injection of atrial myocardial extract in rats," *Journal of the American Society of Nephrology*, vol. 12, no. 2, pp. 403–409, 1981.
- [5] T. Sudoh, N. Minamino, K. Kangawa, and H. Matsuo, "C-Type natriuretic peptide (CNP): a new member of natriuretic peptide family identified in porcine brain," *Biochemical and Biophysical Research Communications*, vol. 168, no. 2, pp. 863–870, 1990.
- [6] S. Doroudgar and C. C. Glembotski, "The cardiokine story unfolds: Ischemic stress-induced protein secretion in the heart," *Trends in Molecular Medicine*, vol. 17, no. 4, pp. 207–214, 2011.
- [7] A. Planavila, J. Fernández-Solà, and F. Villarroya, "Cardiokines as Modulators of Stress-Induced Cardiac Disorders," *Advances in Protein Chemistry and Structural Biology*, vol. 108, pp. 227–256, 2017.
- [8] M. Shimano, N. Ouchi, and K. Walsh, "Cardiokines: recent progress in elucidating the cardiac secretome," *Circulation*, vol. 126, no. 21, pp. e327–e332, 2012.
- [9] Z. Kobalava, "SP 04-1 the role of natriuretic peptides in the pathogenesis of cardiovascular diseases," *Journal of Hypertension*, vol. 34, Suppl 1, p. e377, 2016, ISH 2016 Abstract Book.
- [10] P. K. Santhekadur, D. P. Kumar, M. Seneshaw, F. Mirshahi, and A. J. Sanyal, "The multifaceted role of natriuretic peptides in metabolic syndrome," *Biomedicine & Pharmacotherapy*, vol. 92, pp. 826–835, 2017.
- [11] T. I. Chang, J. Yang, J. V. Freeman, M. A. Hlatky, and A. S. Go, "Effectiveness of β -blockers in heart failure with left ventricular systolic dysfunction and chronic kidney disease," *Journal of Cardiac Failure*, vol. 19, no. 3, pp. 176–182, 2013.
- [12] J. M. Tonne, J. M. Campbell, A. Cataliotti et al., "Secretion of glycosylated pro-B-type natriuretic peptide from normal cardiomyocytes," *Clinical Chemistry*, vol. 57, no. 6, pp. 864–873, 2011.
- [13] B. K. Huntley, S. M. Sandberg, D. M. Heublein, S. Jenson Sangaralingham, J. C. Burnett, and T. Ichiki, "ProB-type natriuretic peptide-1-108 processing and degradation in human heart failure," *Circulation: Heart Failure*, vol. 8, no. 1, pp. 89–97, 2015.
- [14] B. M. Everett, T. Zeller, R. J. Glynn, P. M. Ridker, and S. Blankenberg, "High-sensitivity cardiac troponin I and B-type natriuretic peptide as predictors of vascular events in primary prevention impact of statin therapy," *Circulation*, vol. 131, no. 21, pp. 1851–1860, 2015.
- [15] D. Godkar, K. Bachu, B. Dave, S. Niranjana, and A. Khanna, "B-type natriuretic peptide (BNP) and ProBNP: Role of emerging markers to guide therapy and determine prognosis in cardiovascular disorders," *American Journal of Therapeutics*, vol. 15, no. 2, pp. 150–156, 2008.

- [16] P. C. Wong, J. Guo, and A. Zhang, "The renal and cardiovascular effects of natriuretic peptides," *Advances in Physiology Education*, vol. 41, no. 2, pp. 179–185, 2017.
- [17] S. A. Hunt, W. T. Abraham, M. H. Chin et al., "ACC/AHA 2005 guideline update for the diagnosis and management of chronic heart failure in the adult: a report of the american college of cardiology/american heart association task force on practice guidelines (writing committee to update the 2001 guidelines for the evaluation and management of heart failure): developed in collaboration with the american college of chest physicians and the international society for heart and lung transplantation: endorsed by the heart rhythm society," *Circulation*, vol. 112, no. 12, pp. e154–e235, 2005.
- [18] C. Mueller, "Comparison of the diagnostic accuracy of BNP and NT-proBNP in acute and chronic heart failure [7]," *Clinical Chemistry*, vol. 53, no. 9, pp. 1719–1720, 2007.
- [19] A. G. Semenov and A. G. Katrukha, "Analytical Issues with Natriuretic Peptides - has this been Overly Simplified?" *EJIFCC*, vol. 27, no. 3, pp. 189–207, 2016.
- [20] C. P. Y. Chan and T. H. Rainer, "Pathophysiological roles and clinical importance of biomarkers in acute coronary syndrome," *Advances in Clinical Chemistry*, vol. 59, pp. 23–63, 2013.
- [21] A. Halaris, "Inflammation, Heart Disease, and Depression," *Current Psychiatry Reports*, vol. 15, no. 10, p. 400, 2013.
- [22] N. G. Frangogiannis, "Regulation of the inflammatory response in cardiac repair," *Circulation Research*, vol. 110, no. 1, pp. 159–173, 2012.
- [23] M. M. Ciccone, F. Cortese, M. Gesualdo et al., "A novel cardiac bio-marker: ST2: a review," *Molecules*, vol. 18, no. 12, pp. 15314–15328, 2013.
- [24] E. O. Weinberg, M. Shimpo, G. W. De Keulenaer et al., "Expression and regulation of ST2, an interleukin-1 receptor family member, in cardiomyocytes and myocardial infarction," *Circulation*, vol. 106, no. 23, pp. 2961–2966, 2002.
- [25] A. Lax, J. Sanchez-Mas, M. C. Asensio-Lopez et al., "Mineralocorticoid receptor antagonists modulate galectin-3 and interleukin-33/ST2 signaling in left ventricular systolic dysfunction after acute myocardial infarction," *JACC: Heart Failure*, vol. 3, no. 1, pp. 50–58, 2015.
- [26] R. R. Sun, L. Lu, M. Liu et al., "Biomarkers and heart disease," *European Review for Medical and Pharmacological Sciences*, vol. 18, no. 19, pp. 2927–2935, 2014.
- [27] C. W. Yancy, M. Jessup, B. Bozkurt et al., "ACC/AHA guideline for the management of heart failure: a report of the american college of cardiology foundation/american heart association task force on practice guidelines," *Journal of the American College of Cardiology*, vol. 62, no. 16, pp. e147–e239, 2013.
- [28] W. M. Ansari, S. E. Humphries, A. K. Naveed, O. J. Khan, D. A. Khan, and E. H. Khattak, "Effect of Coronary Artery Disease risk SNPs on serum cytokine levels and cytokine imbalance in Premature Coronary Artery Disease," *Cytokine*, 2017.
- [29] X. Tu, S. Nie, Y. Liao et al., "The IL-33-ST2L pathway is associated with coronary artery disease in a Chinese Han population," *American Journal of Human Genetics*, vol. 93, no. 4, pp. 652–660, 2013.
- [30] H. O. Hambrock, B. Kaufmann, S. Müller et al., "Structural characterization of TSC-36/Flik: Analysis of two charge isoforms," *The Journal of Biological Chemistry*, vol. 279, no. 12, pp. 11727–11735, 2004.
- [31] A. D. Anastasilakis, D. Koulaxis, N. Kefala et al., "Circulating irisin levels are lower in patients with either stable coronary artery disease (CAD) or myocardial infarction (MI) versus healthy controls, whereas follistatin and activin A levels are higher and can discriminate MI from CAD with similar to CK-MB accuracy," *Metabolism - Clinical and Experimental*, vol. 73, pp. 1–8, 2017.
- [32] M. Shimano, N. Ouchi, K. Nakamura et al., "Cardiac myocyte follistatin-like 1 functions to attenuate hypertrophy following pressure overload," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 43, pp. E899–E906, 2011.
- [33] S. Maruyama, K. Nakamura, K. N. Papanicolaou et al., "Follistatin-like 1 promotes cardiac fibroblast activation and protects the heart from rupture," *EMBO Molecular Medicine*, vol. 8, no. 8, pp. 949–966, 2016.
- [34] K. Wei, V. Serpooshan, C. Hurtado et al., "Epicardial FSTL1 reconstitution regenerates the adult mammalian heart," *Nature*, vol. 525, no. 7570, pp. 479–485, 2015.
- [35] A.-K. Alteköester and R. P. Harvey, "Bioengineered FSTL1 Patches Restore Cardiac Function Following Myocardial Infarction," *Trends in Molecular Medicine*, vol. 21, no. 12, pp. 731–733, 2015.
- [36] A. El-Armouche, N. Ouchi, K. Tanaka et al., "Follistatin-like 1 in chronic systolic heart failure: a marker of left ventricular remodeling," *Circulation: Heart Failure*, vol. 4, no. 5, pp. 621–627, 2011.
- [37] K. Tanaka, M. Valero-Muñoz, R. M. Wilson et al., "Follistatin-Like 1 Regulates Hypertrophy in Heart Failure With Preserved Ejection Fraction," *JACC: Basic to Translational Science*, vol. 1, no. 4, pp. 207–221, 2016.
- [38] Y. Ogura, N. Ouchi, K. Ohashi et al., "Therapeutic impact of follistatin-like 1 on myocardial ischemic injury in preclinical models," *Circulation*, vol. 126, no. 14, pp. 1728–1738, 2012.
- [39] Y. Lü, J. H. Liu, L. K. Zhang et al., "Fibroblast growth factor 21 as a possible endogenous factor inhibits apoptosis in cardiac endothelial cells," *Chinese Medical Journal*, vol. 123, no. 23, pp. 3417–3421, 2010.
- [40] X.-L. Lin, X.-L. He, J.-F. Zeng et al., "FGF21 increases cholesterol efflux by upregulating ABCA1 through the ERK1/2-PPAR γ -LXR α pathway in THP1 macrophage-derived foam cells," *DNA and Cell Biology*, vol. 33, no. 8, pp. 514–521, 2014.
- [41] J. Yie, W. Wang, L. Deng et al., "Understanding the physical interactions in the FGF21/FGFR/ β -Klotho complex: structural requirements and implications in FGF21 signaling," *Chemical Biology & Drug Design*, vol. 79, no. 4, pp. 398–410, 2012.
- [42] J. Fernández-Solà and A. P. Porta, "New treatment strategies for alcohol-induced heart damage," *International Journal of Molecular Sciences*, vol. 17, no. 10, article no. 1651, 2016.
- [43] A. Planavila, I. Redondo, E. Hondares et al., "Fibroblast growth factor 21 protects against cardiac hypertrophy in mice," *Nature Communications*, vol. 4, article 2019, 2013.
- [44] A. Planavila, I. Redondo-Angulo, F. Ribas et al., "Fibroblast growth factor 21 protects the heart from oxidative stress," *Cardiovascular Research*, vol. 106, no. 1, pp. 19–31, 2015.
- [45] V. Patel, R. Adya, J. Chen et al., "Novel insights into the cardio-protective effects of FGF21 in lean and obese rat hearts," *PLoS ONE*, vol. 9, no. 2, Article ID e87102, 2014.
- [46] P. Malekar, M. Hagenmueller, A. Anyanwu et al., "Wnt signaling is critical for maladaptive cardiac hypertrophy and accelerates myocardial remodeling," *Hypertension*, vol. 55, no. 4, pp. 939–945, 2010.

- [47] V. A. M. Van De Schans, S. W. M. Van Den Borne, A. E. Strzelecka et al., "Interruption of Wnt signaling attenuates the onset of pressure overload-induced cardiac hypertrophy," *Hypertension*, vol. 49, no. 3, pp. 473–480, 2007.
- [48] K. Marinou, C. Christodoulides, C. Antoniadis, and M. Koutsilieris, "Wnt signaling in cardiovascular physiology," *Trends in Endocrinology & Metabolism*, vol. 23, no. 12, pp. 628–636, 2012.
- [49] E. T. Askevold, P. Aukrust, S. H. Nymo et al., "The cardiokine secreted Frizzled-related protein 3, a modulator of Wnt signalling, in clinical and experimental heart failure," *Journal of Internal Medicine*, vol. 275, no. 6, pp. 621–630, 2014.
- [50] T. Ueland, K. Caidahl, E. T. Askevold, T. Karlsson, M. Hartford, and P. Aukrust, "Secreted Frizzled-Related Protein 3 (sFRP3) in acute coronary syndromes," *International Journal of Cardiology*, vol. 190, no. 1, pp. 217–219, 2015.
- [51] H. Lin, M. Angeli, K. J. Chung, C. Ejimadu, A. R. Rosa, and T. Lee, "sFRP2 activates Wnt/ β -catenin signaling in cardiac fibroblasts: Differential roles in cell growth, energy metabolism, and extracellular matrix remodeling," *American Journal of Physiology-Cell Physiology*, vol. 311, no. 5, pp. C710–C719, 2016.
- [52] S. Martin, H. Lin, C. Ejimadu, and T. Lee, "CALL FOR PAPERS Cell Signaling: Proteins, Pathways and Mechanisms Tissue-nonspecific alkaline phosphatase as a target of sFRP2 in cardiac fibroblasts," *American Journal of Physiology-Cell Physiology*, vol. 309, no. 3, pp. C139–C147, 2015.
- [53] W. He, L. Zhang, A. Ni et al., "Exogenously administered secreted frizzled related protein 2 (Sfrp2) reduces fibrosis and improves cardiac function in a rat model of myocardial infarction," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 107, no. 49, pp. 21110–21115, 2010.
- [54] M. Mastri, Z. Shah, K. Hsieh et al., "Secreted Frizzled-related protein 2 as a target in antifibrotic therapeutic intervention," *American Journal of Physiology-Cell Physiology*, vol. 306, no. 6, pp. C531–C539, 2014.
- [55] N. H. Dayawansa, X.-M. Gao, D. A. White, A. M. Dart, and X.-J. Du, "Role of MIF in myocardial ischaemia and infarction: Insight from recent clinical and experimental findings," *Clinical Science*, vol. 127, no. 3, pp. 149–161, 2014.
- [56] P. V. Tilstam, D. Qi, L. Leng, L. Young, and R. Bucala, "MIF family cytokines in cardiovascular diseases and prospects for precision-based therapeutics," *Expert Opinion on Therapeutic Targets*, vol. 21, no. 7, pp. 671–683, 2017.
- [57] W. Chan, D. A. White, X. Wang et al., "Macrophage Migration Inhibitory Factor for the Early Prediction of Infarct Size," *Journal of the American Heart Association*, vol. 2, no. 5, p. e000226, 2013.
- [58] J.-Y. Luo, R. Xu, X.-M. Li et al., "MIF Gene Polymorphism rs755622 Is Associated With Coronary Artery Disease and Severity of Coronary Lesions in a Chinese Kazakh Population: A Case-Control Study," *Medicine*, vol. 95, no. 4, p. e2617, 2016.
- [59] D. A. White, Y. Su, P. Kanellakis et al., "Differential roles of cardiac and leukocyte derived macrophage migration inhibitory factor in inflammatory responses and cardiac remodelling post myocardial infarction," *Journal of Molecular and Cellular Cardiology*, vol. 69, pp. 32–42, 2014.
- [60] A. Kirabo, S. Ryzhov, M. Gupte et al., "Neuregulin-1 β induces proliferation, survival and paracrine signaling in normal human cardiac ventricular fibroblasts," *Journal of Molecular and Cellular Cardiology*, vol. 105, pp. 59–69, 2017.
- [61] N. Hedhli, Q. Huang, A. Kalinowski et al., "Endothelium-derived neuregulin protects the heart against ischemic injury," *Circulation*, vol. 123, no. 20, pp. 2254–2262, 2011.
- [62] J. D. Gardner, "Neuregulin-1 β as a potential therapeutic for targeting fibroblasts in heart disease," *Journal of Molecular and Cellular Cardiology*, vol. 112, pp. 132–133, 2017.
- [63] K. Krzeminski, "The Role of Adrenomedullin in Cardiovascular Response to Exercise - A Review," *Journal of Human Kinetics*, vol. 53, no. 1, pp. 127–142, 2016.
- [64] B. M. Y. Cheung and F. Tang, "Adrenomedullin: exciting new horizons," *Recent Patents on Endocrine, Metabolic & Immune Drug Discovery*, vol. 6, no. 1, pp. 4–17, 2012.
- [65] F. Beygui, P. S. Wild, T. Zeller et al., "Adrenomedullin and arterial stiffness integrative approach combining monocyte adm expression, plasma MR-Pro-ADM, and genome-wide association study," *Circulation: Cardiovascular Genetics*, vol. 7, no. 5, pp. 634–641, 2014.
- [66] H. K. Wong, T. T. Cheung, and B. M. Cheung, "Adrenomedullin and cardiovascular diseases," *Journal of the Royal Society of Medicine Cardiovascular Disease*, vol. 1, no. 5, 2012.
- [67] H. Nishida, T. Horio, Y. Suzuki et al., "Plasma adrenomedullin as an independent predictor of future cardiovascular events in high-risk patients: Comparison with C-reactive protein and adiponectin," *Peptides*, vol. 29, no. 4, pp. 599–605, 2008.
- [68] H. Tolppanen, M. Rivas-Lasarte, J. Lassus et al., "Adrenomedullin: a marker of impaired hemodynamics, organ dysfunction, and poor prognosis in cardiogenic shock," *Annals of Intensive Care*, vol. 7, no. 1, article no. 6, 2017.
- [69] I. T. Klip, A. A. Voors, S. D. Anker et al., "Prognostic value of mid-regional pro-adrenomedullin in patients with heart failure after an acute myocardial infarction," *Heart*, vol. 97, no. 11, pp. 892–898, 2011.
- [70] R. J. A. Frost and S. Engelhardt, "A secretion trap screen in yeast identifies protease inhibitor 16 as a novel antihypertrophic protein secreted from the heart," *Circulation*, vol. 116, no. 16, pp. 1768–1775, 2007.
- [71] G. G. J. Hazell, A. M. G. Peachey, J. E. Teasdale et al., "PI16 is a shear stress and inflammation-regulated inhibitor of MMP2," *Scientific Reports*, vol. 6, Article ID 39553, 2016.
- [72] A. Tadimallam, P. J. Belmont, D. J. Thuerauf et al., "Mesencephalic astrocyte-derived neurotrophic factor is an ischemia-inducible secreted endoplasmic reticulum stress response protein in the heart," *Circulation Research*, vol. 103, no. 11, pp. 1249–1258, 2008.
- [73] C. C. Glembotski, D. J. Thuerauf, C. Huang, J. A. Vekich, R. A. Gottlieb, and S. Doroudgar, "Mesencephalic astrocyte-derived neurotrophic factor protects the heart from ischemic damage and is selectively secreted upon sarco/endoplasmic reticulum calcium depletion," *The Journal of Biological Chemistry*, vol. 287, no. 31, pp. 25893–25904, 2012.
- [74] C. C. Glembotski, "Functions for the cardiomyokine, MANF, in cardioprotection, hypertrophy and heart failure," *Journal of Molecular and Cellular Cardiology*, vol. 51, no. 4, pp. 512–517, 2011.
- [75] H. Liu, C. Yu, H. Yu et al., "Cerebral dopamine neurotrophic factor protects H9c2 cardiomyocytes from apoptosis," *Herz*, pp. 1–6, 2017.
- [76] Y. Joki, K. Ohashi, D. Yuasa et al., "Neuron-derived neurotrophic factor ameliorates adverse cardiac remodeling after experimental myocardial infarction," *Circulation: Heart Failure*, vol. 8, no. 2, pp. 342–351, 2015.
- [77] E. Pius-Sadowska and B. Machaliński, "BDNF – A key player in cardiovascular system," *Journal of Molecular and Cellular Cardiology*, vol. 110, pp. 54–60, 2017.

- [78] N. Feng, S. Huke, G. Zhu et al., "Constitutive BDNF/TrkB signaling is required for normal cardiac contraction and relaxation," *Proceeding of the National Academy of Sciences of the United States of America*, vol. 112, no. 6, pp. 1880–1885, 2015.
- [79] T. Brandenburger, H. Grievink, N. Heinen et al., "Effects of remote ischemic preconditioning and myocardial ischemia on microRNA-1 expression in the rat heart in vivo," *Shock (Augusta, Ga.)*, vol. 42, no. 3, pp. 234–238, 2014.
- [80] A. Anastasia, K. Deinhardt, S. Wang et al., "Trkb signaling in pericytes is required for cardiac microvessel stabilization," *PLoS ONE*, vol. 9, no. 1, Article ID e87406, 2014.
- [81] Q. Liu and M. T. T. Wong-Riley, "Postnatal development of brain-derived neurotrophic factor (BDNF) and tyrosine protein kinase B (TrkB) receptor immunoreactivity in multiple brain stem respiratory-related nuclei of the rat," *Journal of Comparative Neurology*, vol. 521, no. 1, pp. 109–129, 2013.
- [82] M. Shen, J. Morton, S. T. Davidge, and Z. Kassiri, "Loss of smooth muscle cell disintegrin and metalloproteinase 17 transiently suppresses angiotensin II-induced hypertension and end-organ damage," *Journal of Molecular and Cellular Cardiology*, vol. 103, pp. 11–21, 2017.
- [83] C. Chen, R. Li, R. S. Ross, and A. M. Manso, "Integrins and integrin-related proteins in cardiac fibrosis," *Journal of Molecular and Cellular Cardiology*, vol. 93, pp. 162–174, 2016.
- [84] Y.-H. Zhang, H.-Y. Sun, K.-H. Chen et al., "Evidence for functional expression of TRPM7 channels in human atrial myocytes," *Basic Research in Cardiology*, vol. 107, no. 5, article no. 282, 2012.
- [85] M. George, A. Vijayakumar, S. B. Dhanesh, J. James, and K. Shivakumar, "Molecular basis and functional significance of Angiotensin II-induced increase in Discoidin Domain Receptor 2 gene expression in cardiac fibroblasts," *Journal of Molecular and Cellular Cardiology*, vol. 90, pp. 59–69, 2016.
- [86] H. Xie, R. Tubbs, and B. Yang, "Detection of MGMT promoter methylation in glioblastoma using pyrosequencing," *International Journal of Clinical and Experimental Pathology*, vol. 8, no. 9, pp. 10216–10227, 2015.
- [87] Y. S. Choi, A. B. M. de Mattos, D. Shao et al., "Preservation of myocardial fatty acid oxidation prevents diastolic dysfunction in mice subjected to angiotensin II infusion," *Journal of Molecular and Cellular Cardiology*, vol. 100, pp. 64–71, 2016.
- [88] J. Pöss, C. Ukena, I. Kindermann et al., "Angiotensin-2 and outcome in patients with acute decompensated heart failure," *Clinical Research in Cardiology*, vol. 104, no. 5, pp. 380–387, 2015.
- [89] S. Chen, L. Guo, B. Chen, L. Sun, and M. Cui, "Association of serum angiotensin-1, angiotensin-2 and angiotensin-2 to angiotensin-1 ratio with heart failure in patients with acute myocardial infarction," *Experimental and Therapeutic Medicine*, vol. 5, no. 3, pp. 937–941, 2013.
- [90] A. Lukasz, G. Beutel, P. Kumpers et al., "Angiotensin-2 in Adults with Congenital Heart Disease and Heart Failure," *PLoS ONE*, vol. 8, no. 6, Article ID e66861, 2013.
- [91] J. Lu, F. Liu, F. Chen et al., "Amlodipine and atorvastatin improve ventricular hypertrophy and diastolic function via inhibiting TNF- α , IL-1 β and NF- κ B inflammatory cytokine networks in elderly spontaneously hypertensive rats," *Biomedicine & Pharmacotherapy*, vol. 83, pp. 330–339, 2016.
- [92] S. Toldo, E. Mezzaroma, B. W. Van Tassel et al., "Interleukin-1 β blockade improves cardiac remodeling after myocardial infarction without interrupting the inflammasome in the mouse," *Experimental Physiology*, vol. 98, no. 3, pp. 734–745, 2013.
- [93] A. B. Gustafsson and L. L. Brunton, "Attenuation of cAMP accumulation in adult rat cardiac fibroblasts by IL-1 β and NO: Role of cGMP-stimulated PDE2," *American Journal of Physiology-Cell Physiology*, vol. 283, no. 2, pp. C463–C471, 2002.
- [94] Å. B. Gustafsson and L. L. Brunton, " β -Adrenergic stimulation of rat cardiac fibroblasts enhances induction of nitric-oxide synthase by interleukin-1 β via message stabilization," *Molecular Pharmacology*, vol. 58, no. 6, pp. 1470–1478, 2000.
- [95] E. T. Askevold, L. Gullestad, C. P. Dahl, A. Yndestad, T. Ueland, and P. Aukrust, "Interleukin-6 signaling, soluble glycoprotein 130, and inflammation in heart failure," *Current Heart Failure Reports*, vol. 11, no. 2, pp. 146–155, 2014.
- [96] H. Schuett, R. Oestreich, G. H. Waetzig et al., "Transsignaling of interleukin-6 crucially contributes to atherosclerosis in mice," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 32, no. 2, pp. 281–290, 2012.
- [97] L. De Gennaro, N. D. Brunetti, D. Montrone, F. De Rosa, A. Cuculo, and M. Di Biase, "Subacute inflammatory activation in subjects with acute coronary syndrome and left ventricular dysfunction," *Inflammation*, vol. 35, no. 1, pp. 363–370, 2012.
- [98] O. M. Bhat, P. U. Kumar, N. V. Giridharan, D. Kaul, M. J. M. Kumar, and V. Dhawan, "Interleukin-18-induced atherosclerosis involves CD36 and NF- κ B crosstalk in Apo E-/- mice," *Journal of Cardiology*, vol. 66, no. 1, pp. 28–35, 2015.
- [99] T. B. Opstad, H. Arnesen, A. Å. Pettersen, and I. Seljeflot, "Combined Elevated Levels of the Proinflammatory Cytokines IL-18 and IL-12 Are Associated with Clinical Events in Patients with Coronary Artery Disease: An Observational Study," *Metabolic Syndrome and Related Disorders*, vol. 14, no. 5, pp. 242–248, 2016.
- [100] K. Y. Park and T. H. Heo, "Critical role of TNF inhibition in combination therapy for elderly mice with atherosclerosis," *Cardiovasc Ther*, 2017.
- [101] V. N. Nikolic, T. Jevtovic-Stoimenov, D. Stokanovic et al., "An inverse correlation between TNF alpha serum levels and heart rate variability in patients with heart failure," *Journal of Cardiology*, vol. 62, no. 1, pp. 37–43, 2013.
- [102] A. Savic-Radojevic, S. Radovanovic, T. Pekmezovic et al., "The Role of Serum VCAM-1 and TNF- α as Predictors of Mortality and Morbidity in Patients with Chronic Heart Failure," *Journal of Clinical Laboratory Analysis*, vol. 27, no. 2, pp. 105–112, 2013.
- [103] S. D. Francis Stuart, N. M. De Jesus, M. L. Lindsey, and C. M. Ripplinger, "The crossroads of inflammation, fibrosis, and arrhythmia following myocardial infarction," *Journal of Molecular and Cellular Cardiology*, vol. 91, pp. 114–122, 2016.
- [104] D. J. Duncan, Z. Yang, P. M. Hopkins, D. S. Steele, and S. M. Harrison, "TNF- α and IL-1 β increase Ca²⁺ leak from the sarcoplasmic reticulum and susceptibility to arrhythmia in rat ventricular myocytes," *Cell Calcium*, vol. 47, no. 4, pp. 378–386, 2010.
- [105] D. J. Greensmith and M. Nirmalan, "The effects of tumor necrosis factor-alpha on systolic and diastolic function in rat ventricular myocytes," *Physiological Reports*, vol. 1, no. 4, p. e00093, 2013.
- [106] C.-K. Wu, J.-K. Lee, F.-T. Chiang et al., "Plasma levels of tumor necrosis factor- α and interleukin-6 are associated with diastolic heart failure through downregulation of sarcoplasmic reticulum Ca²⁺ ATPase," *Critical Care Medicine*, vol. 39, no. 5, pp. 984–992, 2011.
- [107] J. Wang, H. Wang, Y. Zhang, H. Gao, S. Nattel, and Z. Wang, "Impairment of HERG K⁺ channel function by tumor necrosis

- factor- α : role of reactive oxygen species as a mediator," *The Journal of Biological Chemistry*, vol. 279, no. 14, pp. 13289–13292, 2004.
- [108] S. E. Sawaya, Y. S. Rajawat, T. G. Rami et al., "Downregulation of connexin40 and increased prevalence of atrial arrhythmias in transgenic mice with cardiac-restricted overexpression of tumor necrosis factor," *American Journal of Physiology-Heart and Circulatory Physiology*, vol. 292, no. 3, pp. H1561–H1567, 2007.
- [109] M. Richter, H.-J. Lautze, T. Walther, T. Braun, S. Kostin, and T. Kubin, "The failing heart is a major source of circulating FGF23 via oncostatin M receptor activation," *The Journal of Heart and Lung Transplantation*, vol. 34, no. 9, article no. 6047, pp. 1211–1214, 2015.
- [110] H. Hao, X. Li, Q. Li et al., "FGF23 promotes myocardial fibrosis in mice through activation of β -catenin," *Oncotarget*, vol. 7, no. 40, pp. 64649–64664, 2016.
- [111] C. Faul, A. P. Amaral, B. Oskouei et al., "FGF23 induces left ventricular hypertrophy," *The Journal of Clinical Investigation*, vol. 121, no. 11, pp. 4393–4408, 2011.
- [112] B. Kestenbaum, M. C. Sachs, A. N. Hoofnagle et al., "Fibroblast growth factor-23 and cardiovascular disease in the general population: the multi-ethnic study of atherosclerosis," *Circulation: Heart Failure*, vol. 7, no. 3, pp. 409–417, 2014.
- [113] R. Jimbo, F. Kawakami-Mori, S. Mu et al., "Fibroblast growth factor 23 accelerates phosphate-induced vascular calcification in the absence of Klotho deficiency," *Kidney International*, vol. 85, no. 5, pp. 1103–1111, 2014.
- [114] V. Olsen, L. E. Rohde, L. Beck-da-Silva et al., "QRS widening rates and genetic polymorphisms of matrix metalloproteinases in a cohort of patients with chronic heart failure," *Canadian Journal of Cardiology*, vol. 30, no. 3, pp. 345–351, 2014.
- [115] M. L. Lindsey and R. Zamilpa, "Temporal and spatial expression of matrix metalloproteinases and tissue inhibitors of metalloproteinases following myocardial infarction," *Cardiovascular Therapeutics*, vol. 30, no. 1, pp. 31–41, 2012.
- [116] M. Quttainah, R. Al-Hejailan, S. Saleh et al., "Progression of matrixin and cardiokine expression patterns in an ovine model of heart failure and recovery," *International Journal of Cardiology*, vol. 186, pp. 77–89, 2015.
- [117] A. Mishra, A. Srivastava, T. Mittal, N. Garg, and B. Mittal, "Association of matrix metalloproteinases (MMP2, MMP7 and MMP9) genetic variants with left ventricular dysfunction in coronary artery disease patients," *Clinica Chimica Acta*, vol. 413, no. 19–20, pp. 1668–1674, 2012.
- [118] J. Medamana, R. A. Clark, and J. Butler, "Platelet-derived growth factor in heart failure," *Handbook of Experimental Pharmacology*, vol. 243, pp. 355–369, 2017.
- [119] D. Yang, J. Yuan, G. Liu et al., "Angiotensin receptor blockers and statins could alleviate atrial fibrosis via regulating platelet-derived growth factor/Rac1/nuclear factor-kappa B axis," *International Journal of Medical Sciences*, vol. 10, no. 7, pp. 812–824, 2013.
- [120] W. Zhao, T. Zhao, V. Huang, Y. Chen, R. A. Ahokas, and Y. Sun, "Platelet-derived growth factor involvement in myocardial remodeling following infarction," *Journal of Molecular and Cellular Cardiology*, vol. 51, no. 5, pp. 830–838, 2011.
- [121] J. Hye Kim, S. Gyu Park, W.-K. Kim, S. U. Song, and J.-H. Sung, "Functional regulation of adipose-derived stem cells by PDGF-D," *Stem Cells*, vol. 33, no. 2, pp. 542–556, 2015.
- [122] Q. Wu, X. Hou, J. Xia et al., "Emerging roles of PDGF-D in EMT progression during tumorigenesis," *Cancer Treatment Reviews*, vol. 39, no. 6, pp. 640–646, 2013.
- [123] M. Christensen, A. J. Najj, M. Snyder, L. S. Movilla, and H.-R. C. Kim, "A critical role of the PTEN/PDGF signaling network for the regulation of radiosensitivity in adenocarcinoma of the prostate," *International Journal of Radiation Oncology • Biology • Physics*, vol. 88, no. 1, pp. 151–158, 2014.
- [124] B. J. He, M.-L. A. Joiner, M. V. Singh et al., "Oxidation of CaMKII determines the cardiotoxic effects of aldosterone," *Nature Medicine*, vol. 17, no. 12, pp. 1610–1618, 2011.
- [125] T. Kempf, A. Zarbock, C. Widera et al., "GDF-15 is an inhibitor of leukocyte integrin activation required for survival after myocardial infarction in mice," *Nature Medicine*, vol. 17, no. 5, pp. 581–588, 2011.
- [126] P. P. Rainer, S. Hao, D. Vanhoutte et al., "Cardiomyocyte-specific transforming growth factor β suppression blocks neutrophil infiltration, augments multiple cytoprotective cascades, and reduces early mortality after myocardial infarction," *Circulation Research*, vol. 114, no. 8, pp. 1246–1257, 2014.
- [127] J. A. Baán, Z. V. Varga, P. Leszek et al., "Myostatin and IGF-I signaling in end-stage human heart failure: a qRT-PCR study," *Journal of Translational Medicine*, vol. 13, no. 1, article no. 1, 2015.
- [128] T. B. Dschietzig, "Myostatin - From the mighty mouse to cardiovascular disease and cachexia," *Clinica Chimica Acta*, vol. 433, pp. 216–224, 2014.
- [129] Y. Oshima, N. Ouchi, M. Shimano et al., "Activin a and follistatin-like 3 determine the susceptibility of heart to ischemic injury," *Circulation*, vol. 120, no. 16, pp. 1606–1615, 2009.
- [130] C. Widera, E. Giannitsis, T. Kempf et al., "Identification of follistatin-like 1 by expression cloning as an activator of the growth differentiation factor 15 gene and a prognostic biomarker in acute coronary syndrome," *Clinical Chemistry*, vol. 58, no. 8, pp. 1233–1241, 2012.
- [131] K. D. Panse, L. E. Felkin, M. M. López-Olañeta et al., "Follistatin-like 3 mediates paracrine fibroblast activation by cardiomyocytes," *Journal of Cardiovascular Translational Research*, vol. 5, no. 6, pp. 814–826, 2012.
- [132] M. Shimano, N. Ouchi, K. Nakamura et al., "Cardiac myocyte-specific ablation of follistatin-like 3 attenuates stress-induced myocardial hypertrophy," *The Journal of Biological Chemistry*, vol. 286, no. 11, pp. 9840–9848, 2011.
- [133] S. Frantz, K. Hu, A. Adamek et al., "Transforming growth factor beta inhibition increases mortality and left ventricular dilatation after myocardial infarction," *Basic Research in Cardiology*, vol. 103, no. 5, pp. 485–492, 2008.
- [134] M. Ikeuchi, H. Tsutsui, T. Shiomi et al., "Inhibition of TGF- β signaling exacerbates early cardiac dysfunction but prevents late remodeling after infarction," *Cardiovascular Research*, vol. 64, no. 3, pp. 526–535, 2004.
- [135] R. J. Akhurst and A. Hata, "Targeting the TGF β signalling pathway in disease," *Nature Reviews Drug Discovery*, vol. 11, no. 10, pp. 790–811, 2012.
- [136] Y. Yang, Y. Li, Z. Ma et al., "A brief glimpse at CTRP3 and CTRP9 in lipid metabolism and cardiovascular protection," *Progress in Lipid Research*, vol. 64, pp. 170–177, 2016.
- [137] M. Appari, A. Breitbart, F. Brandes et al., "Clq-TNF-Related Protein-9 Promotes Cardiac Hypertrophy and Failure," *Circulation Research*, vol. 120, no. 1, pp. 66–77, 2017.
- [138] T. Kambara, K. Ohashi, R. Shibata et al., "CTRP9 protein protects against myocardial injury following ischemia-reperfusion

- through AMP-activated protein kinase (AMPK)-dependent mechanism,” *The Journal of Biological Chemistry*, vol. 287, no. 23, pp. 18965–18973, 2012.
- [139] J. M. Peterson, Z. Wei, M. M. Seldin, M. S. Byerly, S. Aja, and G. W. Wong, “CTRP9 transgenic mice are protected from diet-induced obesity and metabolic dysfunction,” *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, vol. 305, no. 5, pp. R522–R533, 2013.
- [140] H. Su, Y. Yuan, X.-M. Wang et al., “Inhibition of CTRP9, a novel and cardiac-abundantly expressed cell survival molecule, by TNF α -initiated oxidative signaling contributes to exacerbated cardiac injury in diabetic mice,” *Basic Research in Cardiology*, vol. 108, article 315, 2013.
- [141] Y. Yuan, W. B. Lau, H. Su et al., “Clq-TNF-related protein-9, a novel cardioprotective cardiokine, requires proteolytic cleavage to generate a biologically active globular domain isoform,” *American Journal of Physiology-Endocrinology and Metabolism*, vol. 308, no. 10, pp. E891–E898, 2015.
- [142] T. Kambara, R. Shibata, K. Ohashi et al., “Clq/tumor necrosis factor-related protein 9 protects against acute myocardial injury through an adiponectin receptor I-AMPK-dependent mechanism,” *Molecular and Cellular Biology*, vol. 35, no. 12, pp. 2173–2185, 2015.
- [143] K. K. Baskin, C. E. Grueter, C. M. Kusminski et al., “MED13-dependent signaling from the heart confers leanness by enhancing metabolism in adipose tissue and liver,” *EMBO Molecular Medicine*, vol. 6, no. 12, pp. 1610–1621, 2014.
- [144] J. W. S. Jahng, E. Song, and G. Sweeney, “Crosstalk between the heart and peripheral organs in heart failure,” *Experimental & Molecular Medicine*, vol. 48, no. 3, article no. e217, 2016.